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メタデータ	言語: eng 出版者: 公開日: 2019-04-22 キーワード (Ja): キーワード (En): 作成者: Hashihama, Fuminori, Suwa, Shuhei, Kanda, Jota メールアドレス: 所属:
URL	https://oacis.repo.nii.ac.jp/records/1723

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1 Liquid waveguide spectrophotometric measurements of arsenate and particulate arsenic, as
2 well as phosphate and particulate phosphorus, in seawater

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10

11 **Abstract**

12 Sensitive methods for the determination of arsenate and particulate arsenic (PAs), as
13 well as phosphate and particulate phosphorus (PP), in seawater are described. The method for
14 arsenate and phosphate was established by applying automated liquid waveguide
15 spectrophotometry. Because the reaction time for the formation of arsenate-molybdate
16 complex is longer than that for phosphate-molybdate complex, a long Teflon tube submerged
17 in a heating bath was installed in the conventional phosphate flow system. The arsenate was
18 quantified as the difference between absorbances of molybdenum blue dyes with (only
19 phosphate) and without (phosphate + arsenate) arsenate reduction treatment. Contamination
20 was observed in the reagent for arsenate reduction and must be corrected. Linear dynamic
21 ranges up to 1000 nM were confirmed for arsenate and phosphate. The detection limits for
22 arsenate and phosphate were 5 and 4 nM, respectively. Freezing was a reliable sample
23 preservation technique for both arsenate and phosphate. The method for PAs and PP was
24 established by combining conventional persulfate oxidation of PP and the automated liquid
25 waveguide spectrophotometry of arsenate and phosphate. The digestion efficiencies of
26 organic As analogs were >93%. Contamination in the glass fiber filter was negligible. Field
27 tests confirmed that the coefficients of variation (CVs) of 10–19 nM arsenate and 4–151 nM
28 phosphate were 7–20% and 1–25%, respectively, while the CVs of 0.9 nM PAs and 10.2 nM
29 PP were 11 % and 4 %, respectively.

30

31 Keyword: Arsenate, Phosphate, Particulate arsenic, Particulate phosphorus, Liquid waveguide
32 spectrophotometry

33

34 **1 Introduction**

35 Arsenic (As) exists in both dissolved and particulate forms in seawater (Neff 1997;
36 Henke 2009). Dissolved forms include arsenate, arsenite, and organic compounds, such as
37 methylated As. Among them, arsenate is predominant because it is thermodynamically stable
38 in oxygenated seawater. Arsenate has physicochemical properties similar to those of
39 phosphate. Marine microbial communities uptake arsenate, as well as phosphate, which
40 induce toxic effects due to decoupling energy metabolism. This process involves a
41 transformation from dissolved As to particulate As (PAs). To understand this transformation
42 process, interaction between As and phosphorus (P), and toxic effect on marine ecosystem,
43 information on the size and dynamics of dissolved As and PAs, as well as dissolved P and
44 particulate P (PP), are required.

45 Several As species in natural water have been determined using a hydride generation
46 method (Andreae 1977; Cutter et al. 1991; Hasegawa et al. 1994), which can be used to detect
47 nanomolar concentrations of As species (Cutter et al. 2001; Cutter and Cutter 2006).
48 Although this method has been widely applied to the study of As biogeochemistry in oceanic
49 water, its application to As and P interaction study is extremely limited because additional
50 measurements of P species are required. A molybdenum blue spectrophotometric method for
51 the determination of arsenate in natural water has also been proposed (Johnson 1971). In this
52 method, arsenate was quantified as the difference between absorbances of the molybdenum
53 blue dyes, with and without arsenate reduction treatment. This method enabled the
54 determination of both arsenate and phosphate, and was useful in biogeochemical studies on
55 As and P, and their interactions. However, there is little application of the molybdenum blue
56 method to oceanic water. Karl and Tien (1992) quantified arsenate concentration in oceanic
57 water using the molybdenum blue method to examine its interference in phosphate
58 measurement. They adopted a magnesium-induced coprecipitation (MAGIC) procedure

59 before spectrophotometric analysis. This procedure concentrated both phosphate and arsenate,
60 and subsequent spectrophotometric analysis detected them at nanomolar concentrations.

61 Aside from the MAGIC method, sensitive phosphate measurements using 100–250 cm
62 path length liquid waveguide capillary cells (LWCCs) were established (e.g. Zhang and Chi
63 2002; Hashihama et al. 2009; 2013). The LWCC is composed of quartz tubing with a 550 μm
64 inner diameter, and its outer surface is coated with a low-refractive-index cladding material,
65 Teflon-AF. The sensitive method using the LWCC is automated, has no concentration
66 procedure, and performs almost as well as the MAGIC method (Li and Hansell 2008). Along
67 with phosphate determination, the LWCC method is potentially applicable to the
68 determination of nanomolar arsenate in oceanic water. However, to the best of our knowledge,
69 there has been no report of arsenate determination using the LWCC method.

70 Furthermore, a sensitive method for the determination of PP in oceanic water was
71 recently established by Ehama et al. (2016). This method was developed by combining the
72 persulfate oxidation of filtered particles (Suzumura 2008) with sensitive spectrophotometry
73 using an LWCC. In theory, the persulfate oxidation can digest not only PP, but also PAs
74 (Hasegawa et al. 1994). Because PAs oxidizes to arsenate in this process, the concentration
75 can be determined by measuring the arsenate concentration using the LWCC method.

76 In this study, we established LWCC methods for determining arsenate and PAs, as well
77 as phosphate and PP, by modifying the conventional LWCC methods for phosphate
78 (Hashihama et al. 2009; 2013) and PP (Ehama et al. 2016). As the reaction time was longer
79 for the arsenate-molybdate complex than the phosphate-molybdate complex (Johnson 1971),
80 a long Teflon tube in a heating bath was installed in the reaction path of the conventional
81 LWCC method for phosphate. Furthermore, the contamination of As and P in the reagents
82 and filter during sample processing, effect of sample freezing on arsenate and phosphate
83 determinations, and digestion efficiencies of organic As analogs in the persulfate oxidation

84 were carefully examined. The established methods were applied to the determinations of
85 arsenate and PAs, as well as phosphate and PP, in oceanic waters.

86

87 **2 Experimental**

88 **2.1 Reagents**

89 All reagents in this study were of analytical reagent grade and obtained from Wako Pure
90 Chemical Industries (Osaka, Japan). The pure water used to prepare analytical reagents and
91 stock standard solutions was purified by a reverse osmosis and deionization system
92 (Millipore Auto Pure WEX3 and WR600A, Yamato, Japan), which produced water with a
93 resistance of 18.2 MΩ·cm. All plastic and glassware used to store reagents or samples was
94 cleaned carefully using 2% Merck Extran MA 03, and then rinsed with 0.3 M hydrochloric
95 acid and pure water immediately prior to use.

96 The preparation of analytical reagents for automated analysis was based on that for the
97 automated phosphate analysis, as described by Hansen and Koroleff (1999), with the
98 exception of the ascorbic acid solution (Hashihama et al. 2013), which was prepared by
99 dissolving 0.6 g ascorbic acid in 180 mL pure water and then adding 15 mL acetone and 5
100 mL of 15% sodium dodecyl sulfate solution. Using acetone and 15% sodium dodecyl sulfate
101 effectively eliminated baseline drift.

102 Based on the work of Johnson (1971), a reducing reagent (RR) for arsenate reduction
103 was prepared by slowly mixing 20 mL of 1.75 M sulfuric acid into 40 mL of 14% sodium
104 disulfite solution and subsequently mixing with 40 mL of 1.4% sodium thiosulfate solution.
105 The RR was prepared immediately prior to analysis.

106 The reagents for persulfate oxidation were based on those described by Ehama et al.
107 (2016) after Suzumura (2008). A 0.17 M sodium sulfate (Na_2SO_4) solution was used to
108 remove dissolved As and P from the filter, and 3% potassium persulfate ($\text{K}_2\text{S}_2\text{O}_8$) was
109 prepared for use as a digestion reagent.

110

111 **2.2 Standard, blank, and carrier**

112 Disodium hydrogen arsenate heptahydrate and potassium dihydrogen phosphate were
113 used to prepare 5 and 10 mM stock standard solutions, respectively. For the automated
114 analysis of arsenate and phosphate, working standard solutions were prepared by diluting the
115 stock solution with western North Pacific surface water (WNPS). WNPS was also used as a
116 carrier. Arsenate and phosphate-free seawater (APFS), prepared by the MAGIC procedure
117 (Karl and Tien 1992), was used as a blank.

118 For the automated analysis of PAs and PP-digested samples, working standards dissolved
119 in 0.5% K₂S₂O₈ and 3% sodium chloride (NaCl) solution (0.5% K₂S₂O₈ + 3% NaCl) was
120 prepared by mixing the arsenate and phosphate standards dissolved in 3.6% NaCl with
121 autoclaved 3% K₂S₂O₈ [5:1 (v:v)]. The 3% NaCl was used as a carrier. The 0.5% K₂S₂O₈ +
122 3% NaCl was used as a blank (reagent blank, see below).

123

124 **2.3 Automated analysis of arsenate and phosphate**

125 A gas-segmented continuous flow system (AutoAnalyzer II, Technicon, now Seal
126 Analytical, Hampshire, UK) was used for the automated analysis of arsenate and phosphate
127 (Fig. 1). In the detector position, we installed a 100 cm path length LWCC (LWCC-2100,
128 World Precision Instruments, Sarasota, FL, USA), a tungsten fiber optic light source (L7893,
129 Hamamatsu Photonics, Shizuoka, Japan), and a miniature fiber optic spectrometer (USB4000,
130 Ocean Optics, Dunedin, FL, USA). Absorption spectra of arsenate and phosphate were
131 recorded with the spectrometer.

132 The present analytical system was based on our analytical system for nanomolar
133 phosphate (Hashihama et al. 2009; 2013), except for a heating bath unit comprising a 2 mm
134 i.d. Teflon tubing (AS ONE, Osaka, Japan) in a temperature-controlled water bath (Isotemp
135 2320, Thermo Fisher Scientific, Waltham, MA, USA). The heating bath unit was attached for
136 the complete formation of arsenate-molybdate complex (Johnson 1971). To determine

137 optimal conditions for the formation of the arsenate-molybdate complex, we monitored the
138 absorbance of 100 nM arsenate working standard at reaction temperatures of 25 and 37 °C at
139 reaction times ranging from 35 to 110 min using different lengths of Teflon tubing. These
140 reaction conditions were preliminarily determined based on those of our phosphate system
141 (Hashihama et al. 2009; 2013) and original manual procedure (Johnson 1971).

142 As arsenate and phosphate were quantified from the absorbances of the molybdenum
143 blue dyes both with and without arsenate reduction, samples with and without RR addition
144 were sequentially injected from a sample suction line of the flow system (Fig. 1). The RR
145 was added to the sample at a volume ratio of 1:10 (RR:sample) (Johnson 1971). The sample
146 mixed with RR was left to stand for 15 min prior to injection. To examine contamination in
147 the RR, absorbance of the RR itself was determined by measuring the RR dissolved in APFS
148 (APFS + RR) at a volume ratio of 1:10.

149

150 **2.4 Sample processing for PAs and PP**

151 Sample processing for PAs and PP was based on that described by Ehama et al. (2016)
152 after Suzumura (2008). A pre-combusted, acid-washed glass fiber filter (Whatman GF/F, 2.5
153 cm diameter, Kent, UK) was used to collect particles. Filtration was carried out using
154 aspirator (A-3S, TOKYORIKAKIKAI, Tokyo, Japan) under vacuum at <0.02 MPa. After
155 filtration, the filter was rinsed with 5 mL of 0.17 M Na₂SO₄, then dried and placed into a
156 digestion glass bottle (GL32, Duran, Wertheim/Main, Germany). The particles on the filter
157 were digested with 20 mL of 3% K₂S₂O₈ at 120 °C for 30 min using an autoclave (KTS-2322,
158 ALP, Tokyo, Japan). The bottle was shaken before and after autoclaving. The residue in the
159 digested solution was removed using a 0.45 µm syringe filter (Millex-HV, Millipore,
160 Massachusetts, USA). Because >2% K₂S₂O₈ inhibits color development in the sample, the
161 digested solutions were diluted six-fold with 3.6% NaCl (i.e., the sample was dissolved in

162 0.5% $K_2S_2O_8$ + 3% NaCl). Arsenate and phosphate concentrations in the diluted solution
163 were determined using the LWCC method described above. In the previous LWCC method
164 for PP, pure water was used as a carrier and to prepare blank and matrices of working
165 standards and samples (Ehama et al. 2016). However, in the present LWCC method for PAs
166 and PP, 3% NaCl was used instead of pure water, because our preliminary experiments
167 showed that using pure water disturbed periodic gas-segmented flow of the analytical system
168 with long Teflon tubing.

169 Almost complete digestion of oceanic PP during processing has been reported elsewhere
170 (Suzumura 2008), while the digestion efficiency of PAs in this processing was not reported.
171 The digestion efficiency of PAs was confirmed using two methylated As analogs,
172 methylarsonic acid and cacodylic acid (Table 1), which are commonly presented in oceanic
173 water (Cutter et al. 2001; Cutter and Cutter 2006). These analogs were dissolved in pure
174 water, and a portion of each solution, containing 12 nmol of As, was dispensed into digestion
175 glass bottles. The analog solutions were gently heated to dryness on a hot plate. The dried
176 samples were digested and analyzed as described above. The final As concentration of each
177 sample was 100 nM, and the digestion efficiency was estimated as a proportion of the
178 absorbance of the digested analogs to the absorbance of the 100 nM arsenate standard. To
179 examine the contamination of arsenate in the analogs, undigested samples were also
180 analyzed.

181 The absorbances of the procedural blank (GF/F filter + 0.5% $K_2S_2O_8$ + 3% NaCl) and
182 reagent blank (0.5% $K_2S_2O_8$ + 3% NaCl) were compared to check As and P contamination in
183 the GF/F filter. The procedural blank was prepared by filtering 1 L of pure water and was
184 processed following the digestion procedure outlined.

185

186 **2.5 Field sampling**

187 Seawater samples for arsenate and phosphate were collected in oceanic waters of the
188 tropical and subtropical Pacific Ocean (37.5°N – 30.0°S) during the KH-11-10 cruise (Dec.
189 2011 – Jan. 2012) and KH-12-3 cruise (Jul. – Aug. 2012) of R/V *Hakuho-maru*. Sampling
190 was conducted using Teflon-coated 12 L Niskin-X bottles (General Oceanics, Miami, FL,
191 USA) on a CTD-Carousel system (Sea-Bird Electronics, Bellevue, WA, USA). The sample in
192 the Niskin-X bottle was poured into 30 mL polypropylene tube (Sarstedt, Nümbrecht,
193 Germany). For testing the reproducibility of arsenate and phosphate determinations, five
194 replicate samples were collected at depths of 119 m in the central North Pacific, 100 m in the
195 eastern South Pacific, and 5 and 100 m in the western North Pacific (Table 2). All samples
196 were stored frozen at –20 °C during the cruises, and then thawed and analyzed onshore using
197 the automated analytical system for arsenate and phosphate. To examine the effect of sample
198 freezing on arsenate and phosphate determinations, each sample obtained from depths of 5
199 and 100 m in the western North Pacific were poured into an additional five tubes. These
200 replicate samples were immediately analyzed on board.

201 Seawater samples for testing the reproducibility of PAs and PP determinations were
202 collected at a depth of 10 m in the western North Pacific during the KH-13-7 cruise of R/V
203 *Hakuho-maru* (Table 3). The sample was collected using Teflon-coated 12 L Niskin-X bottles
204 on a CTD-Carousel system, and was poured into three polycarbonate bottles (Thermo
205 Scientific Nalgene, Rochester, NY, USA). Each sample, with a volume of 1.19 L, was filtered
206 using a GF/F filter. The filters were stored at –20 °C until onshore analysis.

207 **3 Results and discussion**

208 **3.1 Arsenate and phosphate determinations**

209 **3.1.1 Absorption spectra**

210 Absorption spectra between 500 and 900 nm were obtained by measuring the 100 nM
211 arsenate and 100 nM phosphate working standards (Fig. 2). Baselines were drawn using the
212 WNPS. The arsenate and phosphate spectra derived from the present LWCC method were
213 similar to those from the original method (Johnson 1971). Due to effective detection of the
214 100 cm LWCC in the range 230 to 730 nm, absorbances for both arsenate and phosphate
215 highly fluctuated at >730 nm. Because the fluctuation slightly extended to the range 710 to
216 730 nm for both arsenate and phosphate, we selected a wavelength of 708 nm for both
217 arsenate and phosphate measurements. This wavelength was also used in our original LWCC
218 method for phosphate (Hashihama et al. 2009; 2013).

219

220 **3.1.2 Optimal conditions for the formation of arsenate-molybdate complex**

221 Color development of arsenate-molybdate complex depended on reaction time and
222 temperature (Fig. 3). In Fig. 3, the absorbance of WNPS at each reaction time was set to zero.
223 At 37 °C, the absorbance of 100 nM arsenate increased with time, reaching a plateau at 95
224 min. The mean absorbance at 95 min was not significantly different from that at 110 min
225 (*t*-test, *p* > 0.05, *n* = 3). At 25 °C, the absorbance of 100 nM arsenate gradually increased, but
226 did not reach the maximum absorbance observed at 37 °C. According to these results, we
227 determined that the optimal reaction time and temperature were 95 min and 37 °C,
228 respectively. These conditions were similar to that of the original method (Johnson 1971).

229

230 **3.1.3 Contamination in reducing reagent**

231 When the absorbance of WNPS was set to zero, the absorbance of APFS + RR and of
232 APFS alone were -0.008 ± 0.003 and -0.014 ± 0.003 , respectively [mean \pm standard
233 deviation (SD), $n = 4$]. The absorbance of APFS + RR was significantly higher than that of
234 APFS (t -test, $p < 0.05$), with a difference of 0.006. This difference was ascribed to any
235 contamination in the RR that increased absorbance (e.g., trace phosphate). Because the
236 contamination resulted in significant overestimation of the phosphate concentration, its
237 absorbance had to be corrected during calculations (see below).

238

239 **3.1.4 Output signals**

240 The typical output signals of WNPS with RR (WNPS + RR), APFS, APFS + RR, 100
241 nM arsenate standard, 100 nM arsenate standard with RR (100 nM arsenate standard + RR),
242 and 100 nM phosphate standard, along with WNPS injections, are shown in Fig. 4. Each
243 suction time was 5 min, which was enough to eliminate the influence of carryover and sample
244 dispersion. Based on sample suction time and the sample line flow rate (1.20 mL min^{-1} , Fig.
245 1), the sample volumes required were estimated to be 6 mL. In the output signals of Fig. 4,
246 WNPS absorbance was set to zero. The absorbance of WNPS + RR (-0.006 ± 0.001 , mean \pm
247 SD, $n = 3$) was slightly lower than that of WNPS. This was ascribed to the reduction of
248 arsenate dissolved in WNPS. The absorbance of the WNPS + RR was not significantly
249 different from that of the 100 nM arsenate standard + RR (t -test, $p > 0.05$, $n = 3$), indicating
250 that arsenate was completely reduced even at the 100 nM level. As mentioned above, the
251 absorbance of APFS + RR was higher than that of APFS. The absorbance of the 100 nM
252 arsenate standard was significantly lower than that of the 100 nM phosphate standard (t -test,
253 $p < 0.05$, $n = 3$). These measurements must be made prior to sample measurement to check
254 arsenate reduction efficiency, contamination in the RR, and the absorbance signals of both
255 arsenate and phosphate.

256

257 **3.1.5 Calibration curves and detection limits**

258 Calibration curves were established using working standards of arsenate and phosphate
259 (Fig. 5). In each curve, WPNS absorbance was set to zero. The linear absorbance response to
260 arsenate and phosphate concentrations up to 1000 nM ($n = 10$) and 40 nM ($n = 5$) were
261 obtained with strong correlations ($r^2 > 0.9971$). Detection limits for arsenate and phosphate
262 were estimated as dividing 3SD of the absorbance of APFS + RR ($n = 7$) by each slope
263 (nM^{-1}) of the calibration curve. The detection limits were 5 and 4 nM for arsenate and
264 phosphate, respectively.

265

266 **3.1.6 Calculations of arsenate and phosphate concentrations in natural samples**

267 The absorbance of APFS was set to zero as a blank when calculating arsenate and
268 phosphate concentrations in natural samples. Arsenate and phosphate concentrations (nM) in
269 natural samples were calculated as follows

$$270 \text{ [Arsenate]} = (\text{ABS}_{\text{sample}} - (\text{ABS}_{\text{sample+RR}} - \text{ABS}_{\text{APFS+RR}}) \times 1.1) / S_{\text{As}} \quad (1)$$

$$271 \text{ [Phosphate]} = (\text{ABS}_{\text{sample+RR}} - \text{ABS}_{\text{APFS+RR}}) \times 1.1 / S_{\text{P}} \quad (2)$$

272 where $\text{ABS}_{\text{sample}}$ is the absorbance of the sample, $\text{ABS}_{\text{sample+RR}}$ is the absorbance of the
273 sample with RR, $\text{ABS}_{\text{APFS+RR}}$ is the absorbance of APFS with RR, 1.1 is the dilution factor
274 with a volume ratio of 11:10 (sample + RR:sample), and S_{As} and S_{P} are the slopes (nM^{-1}) of
275 the calibration curves of arsenate and phosphate, respectively.

276

277 **3.1.7 Effect of sample freezing on arsenate and phosphate determinations**

278 Arsenate concentrations in the fresh samples (immediately analyzed onboard) collected
279 at 5 and 100 m in the western North Pacific were 10 ± 2 and 14 ± 1 (mean \pm SD, $n = 5$),
280 respectively, while those in frozen samples collected at 5 and 100 m in the western North

281 Pacific were 11 ± 1 and 15 ± 1 ($n = 5$), respectively (Table 2). Arsenate concentrations in
282 fresh samples were not significantly different to those in frozen samples (t -test, $p > 0.05$, $n =$
283 5). Phosphate concentrations in the fresh samples at 5 and 100 m in the western North Pacific
284 were 4 ± 1 and 5 ± 1 ($n = 5$), respectively, while those in the frozen samples at 5 and 100 m in
285 the western North Pacific were 4 ± 1 and 6 ± 1 ($n = 5$), respectively. As for arsenate
286 concentrations, phosphate concentrations in fresh samples were not significantly different to
287 those in frozen samples (t -test, $p > 0.05$, $n = 5$). These results indicated that freezing was a
288 reliable method for the preservation of arsenate and phosphate samples.

289

290 **3.1.8 Concentration and reproducibility of the filed sample**

291 Arsenate and phosphate concentrations in the field samples collected at depths of 10 m in
292 the tropical and subtropical Pacific ranged from 8 to 20 nM and from <4 to 471 nM,
293 respectively ($n = 33$). The concentration range of arsenate was fairly uniform and much
294 smaller than that of phosphate. The arsenate concentration level was well consistent with that
295 previously observed at the surface (<10 m) of the tropical and subtropical Pacific (12 – 28
296 nM, Karl and Tien 1992; Cutter and Cutter 2006). The present arsenate measurement is not
297 influenced by other As species, because the concentrations of arsenite and organic As in
298 oceanic waters are generally at picomolar or subnanomolar level (Cutter et al. 2001; Cutter
299 and Cutter 2006) and far below the detection limit (5 nM) of our analytical system. Thus, the
300 present analytical method could measure reliable arsenate concentrations in oceanic waters.

301 In contrast, the phosphate concentrations derived from the present analytical method
302 were compared with those derived from the conventional liquid waveguide
303 spectrophotometry of phosphate (Hashihama et al. 2009; 2013) (Fig. 6). Data from the
304 conventional method were obtained from 10 m at the same stations in the KH-11-10 and
305 KH-12-3 cruises and were previously reported in Sato et al. (2013). The phosphate

306 concentrations derived from two methods were not significantly different from each other (all
307 samples: paired *t*-test, $p > 0.05$, $n = 21$), even in low concentration range (<50 nM samples:
308 paired *t*-test, $p > 0.05$, $n = 9$). The result confirmed that modification of the conventional
309 method did not influence on phosphate determination; for example, there was no
310 decomposition of organic P into phosphate due to long reaction time (95 min) and high
311 reaction temperature (37 °C). Furthermore, the conventional phosphate method had no
312 interference from arsenate and silicic acid (Hashihama et al. 2013). This indicates that, in the
313 present phosphate method, arsenate was completely reduced and the interference from silicic
314 acid was negligible.

315 Mean concentrations of arsenate and phosphate in the replicate samples ranged from 10
316 to 19 nM and from 4 to 151 nM, respectively (Table 2). Although very low concentrations
317 were observed in several samples, the coefficients of variation (CVs) for replicate samples (n
318 = 5) were less than 20% and 25% for arsenate and phosphate, respectively. These data were
319 good quality and the present method is suitable for the measurements of oligotrophic samples.
320 However, there remains a concern for arsenate determination in low-arsenate and
321 high-phosphate environments such as nutrient-rich deep water, because high phosphate
322 concentrations at micromolar levels generally have large measurement errors at nanomolar
323 levels, which accompany large errors of nanomolar arsenate through the calculation. In the
324 low-arsenate and high-phosphate environments, the present method is unsuitable and the
325 arsenate determination should be done using a hydride generation method (Andreae 1977;
326 Cutter et al. 1991; Hasegawa et al. 1994).

327

328 **3.2 PAs and PP determinations**

329 **3.2.1 Digestion efficiencies of organic As analogs**

330 The absorbances of undigested organic As analogs were not significantly different from
331 the blank absorbance (t -test, $p > 0.05$, $n = 3$), indicating that organic As analogs did not
332 contain inorganic arsenate. A comparison of the absorbances of digested organic As analogs
333 and the arsenate standard revealed that the digestion efficiencies of methylarsonic acid and
334 cacodylic acid were $93 \pm 5\%$ and $99 \pm 6\%$ (mean \pm SD, $n = 3$), respectively (Table 1). These
335 results implied that the sample processing in this study could digest PAs in the same manner
336 previously observed for PP (Suzumura 2008).

337

338 **3.2.2 Contamination in glass fiber filter**

339 When the absorbance of 3% NaCl (carrier) was set to zero, the absorbances of the
340 reagent blank and procedural blank were both 0.009 ± 0.001 (mean \pm SD, $n = 3$), with no
341 significant differences (t -test, $p > 0.05$, $n = 3$). Andreae (1999) described the glass fiber filter
342 potentially containing trace As compounds. However, based on our results, the GF/F filter
343 used here did not contain either As or P compounds. Thus, contamination in the GF/F filter
344 was negligible.

345

346 **3.2.3 Calculations of PAs and PP concentrations in natural samples**

347 Absorbance of the reagent blank (0.5% $K_2S_2O_8$ + 3% NaCl) was set to zero when
348 calculating PAs and PP concentrations in natural samples. PAs and PP concentrations (nM) in
349 natural samples were calculated as follows

$$350 \text{ [PAs]} = [\text{Arsenate}_{\text{digested}}] \times 0.02 \times 6 / \text{FV} \quad (3)$$

$$351 \text{ [PP]} = [\text{Phosphate}_{\text{digested}}] \times 0.02 \times 6 / \text{FV} \quad (4)$$

352 where $[\text{Arsenate}_{\text{digested}}]$ and $[\text{Phosphate}_{\text{digested}}]$ are the concentrations (nM) of arsenate and
353 phosphate in the digested solution, respectively, 0.02 is the volume (L) of 3% $K_2S_2O_8$, 6 is
354 the dilution factor with a volume ratio of 6:1 (digested solution with 3.6% NaCl:digested

355 solution), and FV is the filtration volume (L). Calculations of [Arsenate_{digested}] and
356 [Phosphate_{digested}] were followed by Eq. (1) and (2), respectively, except for ABS_{APFS+RR}. In
357 place of the ABS_{APFS+RR}, the absorbance of RR dissolved in the reagent blank was used here.
358 The slopes (nM^{-1}) of the calibration curves (S_{As} and S_P) used here should be derived from the
359 working standards dissolved in 0.5% $\text{K}_2\text{S}_2\text{O}_8$ + 3% NaCl, but slope values were not
360 significantly different between the standard matrices of WNPS and 0.5% $\text{K}_2\text{S}_2\text{O}_8$ + 3% NaCl
361 (t -test, $p > 0.05$, $n = 3$).

362

363 **3.2.4 Concentration and reproducibility of the field sample**

364 PAs and PP concentrations in the samples collected at a depth of 10 m in the western
365 North Pacific were 0.9 ± 0.1 and 10.2 ± 0.4 (mean \pm SD, $n = 3$), respectively (Table 3). To
366 the best of our knowledge, these are the first data for both PAs and PP concentrations in
367 oceanic water, although the data were obtained from single location. The CVs of the PAs and
368 PP samples were 11 and 4%, respectively. The PP concentration with the CV was consistent
369 with those of other oceanic water (Ehama et al. 2016). Indeed, the PAs concentration in
370 oceanic water was very low. If standard spectrophotometry, which generally measures
371 micromolar levels of arsenate, was adopted, 10–1000 fold filtration volumes would be
372 required compared with the nanomolar measurement of arsenate. In the present PAs method,
373 which is a technique for determining nanomolar arsenate, accurate measurements with low
374 CV were possible even at practical filtration volumes (1.19 L).

375 **4 Conclusion**

376 The present study established sensitive analytical methods for the determination of
377 arsenate and PAs, as well as phosphate and PP, in seawater using liquid waveguide
378 spectrophotometry. The arsenate measurement was performed by installing a heating bath
379 unit with long Teflon tubing. Careful assessments of contamination in the reagents and filter,
380 sample preservation, and digestion efficiency enabled accurate measurement of natural
381 samples. The present methods are designed for the determination of arsenate, phosphate, PAs,
382 and PP in oceanic water. By applying this new method to extensive observation in oceanic
383 areas, we can gain a better understanding of oceanic As and P dynamics.

384 **Acknowledgments**

385 We are grateful to the officers, crew members, and members of the scientific party of the
386 R/V *Hakuho-maru* cruises for their cooperation at sea. This work was financially supported
387 by Grant-in-Aid No. 24710004 for Young Scientists (B) from the Japan Society for
388 Promotion of Science and Grant-in Aid No. 24121003 for Scientific Research in Innovative
389 Areas (New Ocean Paradigm on Its Biogeochemistry, Ecosystem and Sustainable Use
390 “NEOPS”) from the Ministry of Education, Culture, Sports, Science and Technology, Japan.

391

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437 natural waters with liquid waveguide. *Environ Sci Technol* 36: 1048–1053

438

439 **Tables**

440 Table 1. Organic As analogs and their digestion efficiencies.

Organic As analog	Formula	Purity (%)	Digestion efficiency \pm SD (%) ($n = 3$)
Methylarsonic acid	CH ₃ AsO ₃	95	93 \pm 5
Cacodylic acid	(CH ₃) ₂ As(O)OH	95	99 \pm 6

441

442 Table 2. Arsenate and phosphate samples collected in oceanic waters and their

443 concentrations.

Sampling location	Date	Latitude	Longitude	Depth (m)	Sample type	Mean concentration \pm SD (nM) [CV %]	
						Arsenate ($n = 5$)	Phosphate ($n = 5$)
Central North Pacific	2011/12/13	23.00 °N	180.00 °	119	Frozen	19 \pm 2 [11]	42 \pm 1 [2]
Eastern South Pacific	2012/1/7	23.00 °S	120.00 °W	100	Frozen	16 \pm 2 [13]	151 \pm 1 [1]
Western North Pacific	2012/7/18	25.00 °N	160.00 °E	5	Fresh	10 \pm 2 [20]	4 \pm 1 [25]
					Frozen	11 \pm 1 [9]	4 \pm 1 [25]
				100	Fresh	14 \pm 1 [7]	5 \pm 1 [20]
					Frozen	15 \pm 1 [7]	6 \pm 1 [17]

444

445 Table 3. Information for filed samples of PAs and PP and their concentrations.

Sampling location	Date	Latitude	Longitude	Depth (m)	Filtered volume (L)	Mean concentration \pm SD (nM) [CV %]	
						PAs ($n = 3$)	PP ($n = 3$)
Western North Pacific	2013/12/16	20.00 °N	160.00 °E	119	1.19	0.9 \pm 0.1 [11]	10.2 \pm 0.4 [4]

446

447 Figure captions

448 Figure 1. Manifold configuration and flow diagram for gas-segmented continuous-flow
449 analysis of arsenate and phosphate with a 100 cm LWCC. The flow rate of each
450 pumping tube is provided to the right of the peristaltic pump.

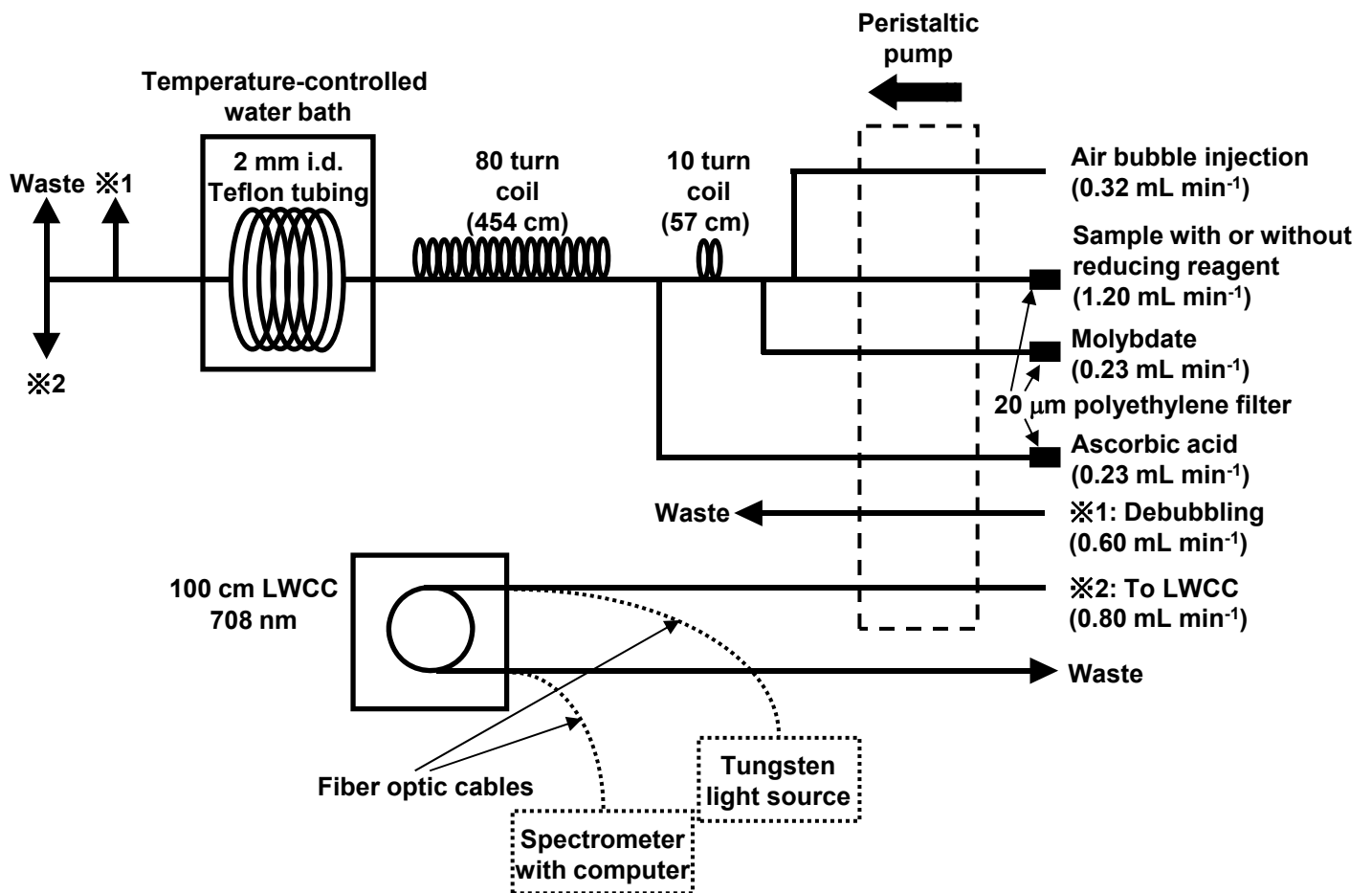
451 Figure 2. Absorption spectra of (a) arsenate-molybdate complex and (b) phosphate-molybdate
452 complex measured in the 100 nM arsenate and 100 nM phosphate working standards,
453 respectively.

454 Figure 3. Absorbance of 100 nM arsenate standard at reaction temperatures 25 °C (open
455 circle) and 37 °C (closed circle) at reaction times ranging from 35 to 110 min. Error
456 bars indicates standard deviations ($n = 3$).

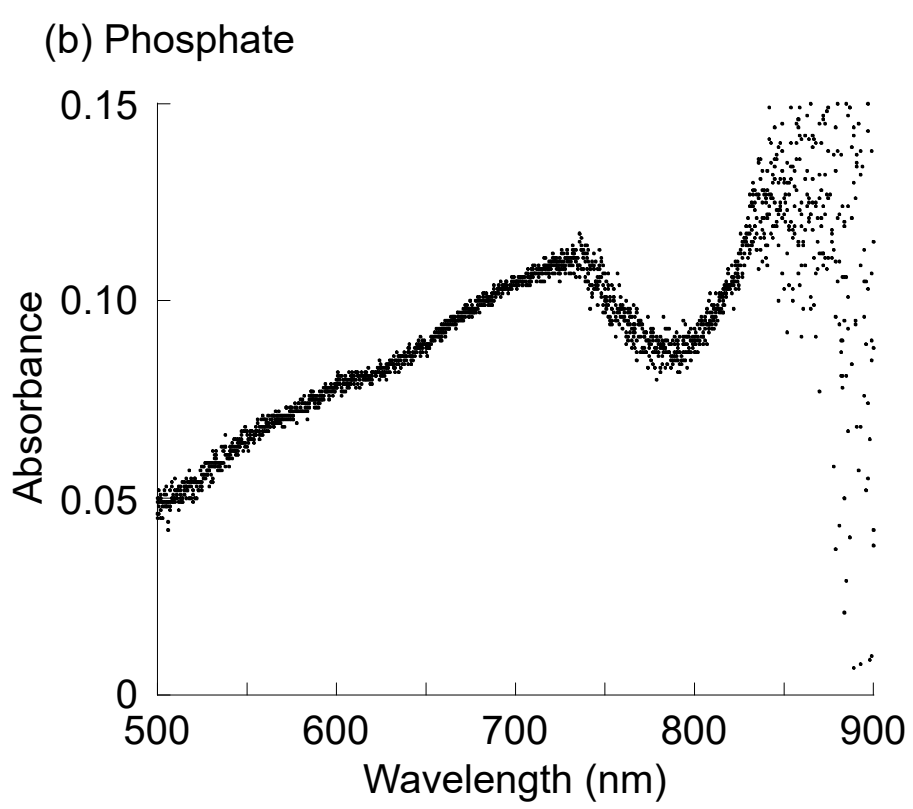
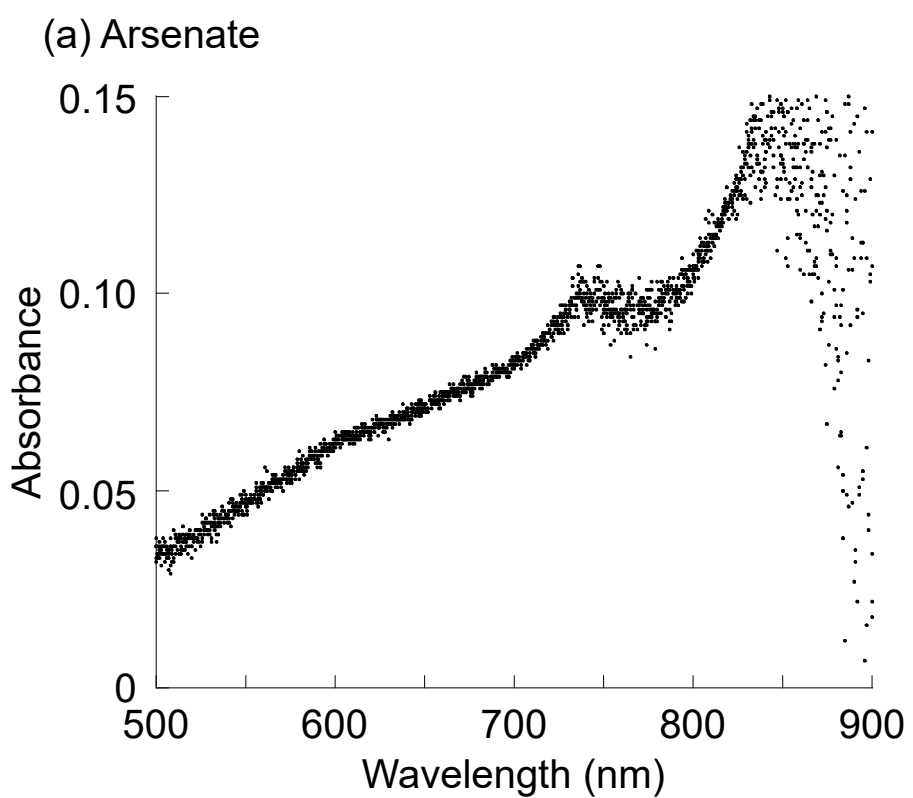
457 Figure 4. Typical output signals of WNPS + RR, APFS, APFS + RR, 100 nM arsenate
458 standard, 100 nM arsenate standard + RR, and 100 nM phosphate standard along with
459 WNPS injections.

460 Figure 5. Calibration curves of arsenate up to (a) 1000 nM and (b) 40 nM and phosphate up to
461 (c) 1000 nM and (d) 40 nM.

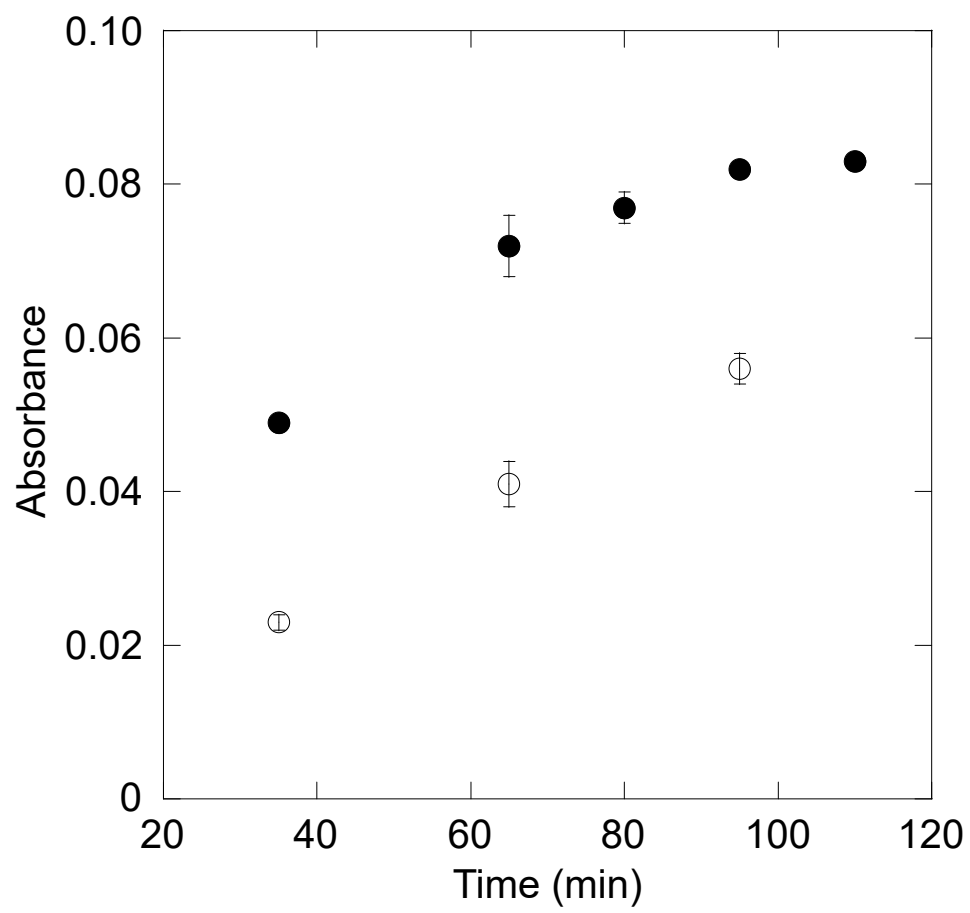
462 Figure 6. Phosphate concentrations of the field samples derived from the present and the
463 conventional analytical methods.



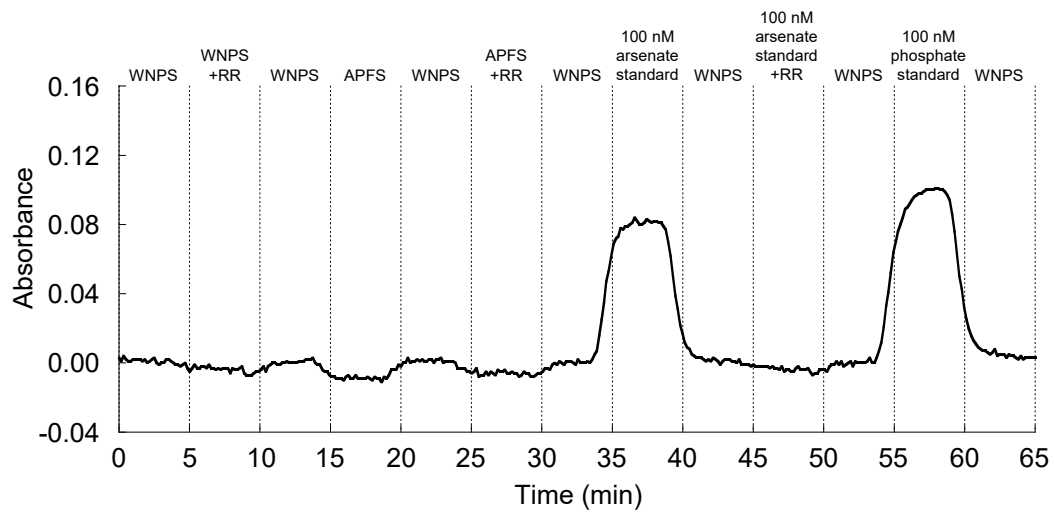
Hashihama et al. Fig. 1



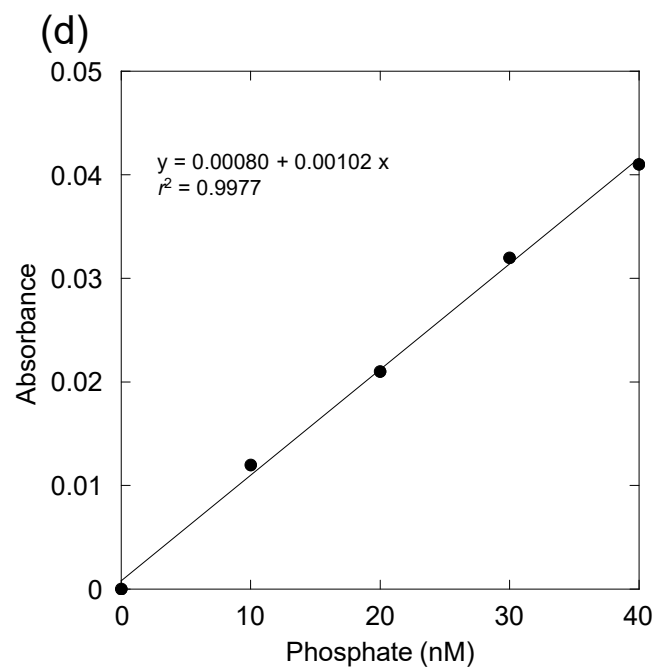
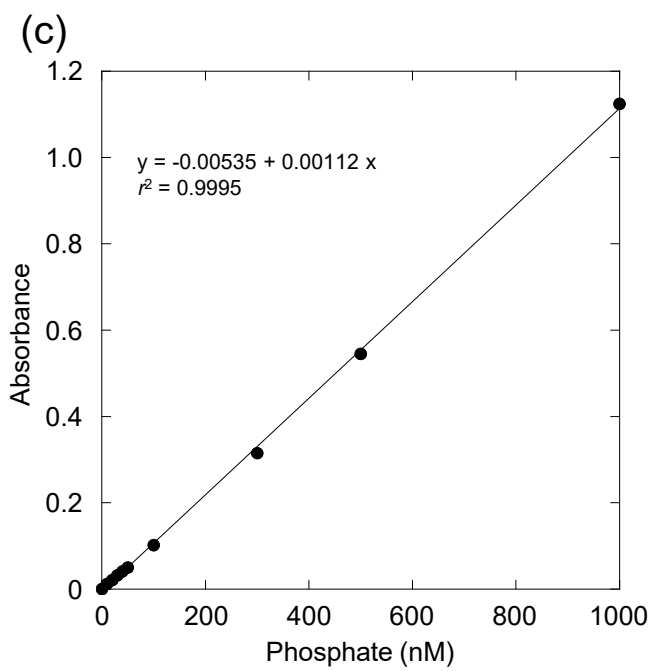
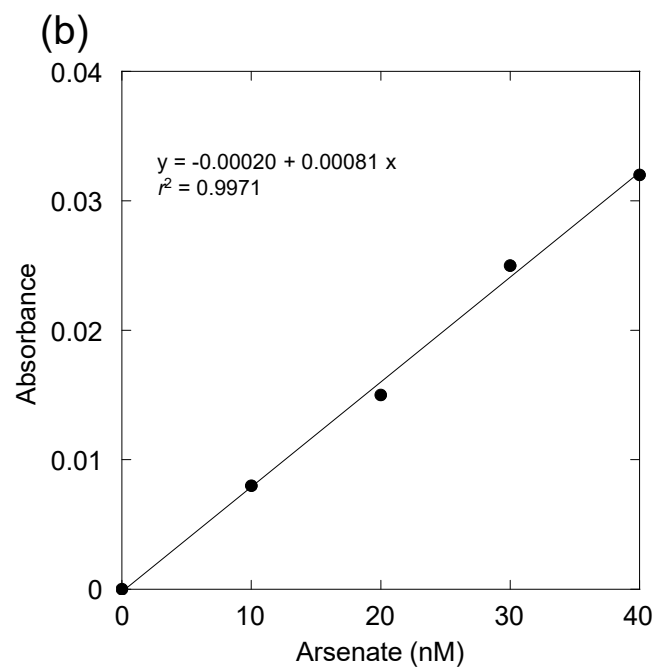
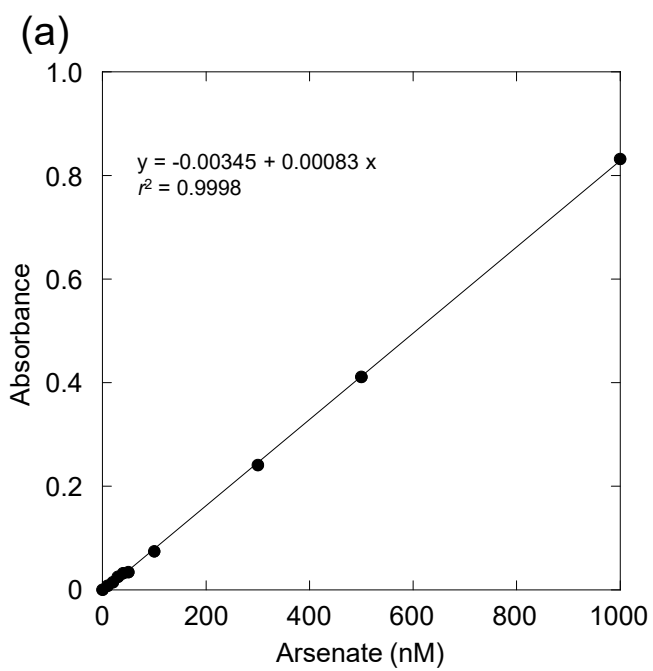
Hashihama et al. Fig. 2



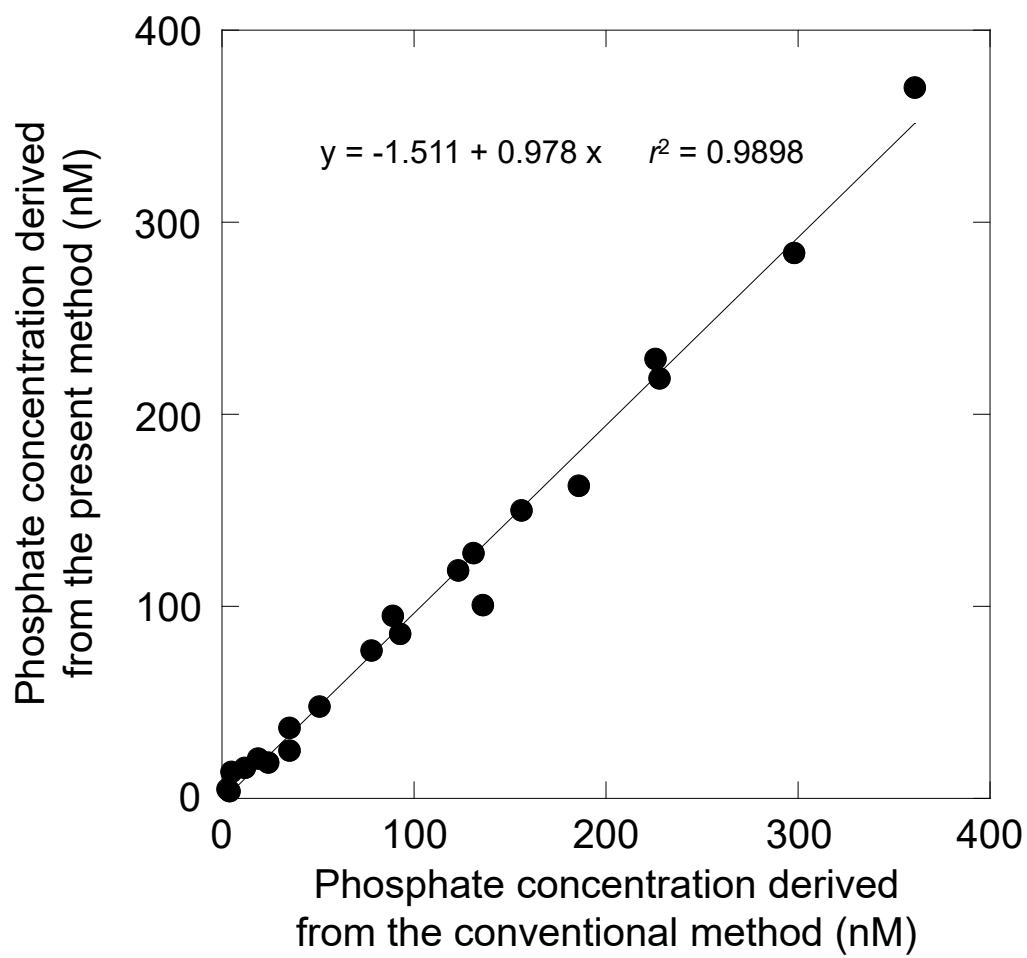
Hashihama et al. Fig. 3



Hashihama et al. Fig. 4



Hashihama et al. Fig. 5



Hashihama et al. Fig. 6